



## DIAGNOSTIC AUTOMATION, INC.

23961 Craftsman Road, Suite E/F, Calabasas, CA 91302

Tel: (818) 591-3030 Fax: (818) 591-8383

[onestep@rapidtest.com](mailto:onestep@rapidtest.com)

[technicalsupport@rapidtest.com](mailto:technicalsupport@rapidtest.com)

[www.rapidtest.com](http://www.rapidtest.com)



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2°C-8°C



Σ=96 tests



Cat # 1801Z

### MICROWELL ELISA IMMUNOGLOBULIN E (IgE) ENZYME IMMUNOASSAY TEST KIT

# IgE

Cat # 1801Z

#### Enzyme Immunoassay for the Quantitative Determination of Immunoglobulin E (IgE) Concentration in Human Serum

#### Intended use

For the quantitative determination of Immunoglobulin E (IgE) concentration in human serum.

#### Introduction

Patients with atopic allergic diseases such as atopic asthma, atopic dermatitis, and hay fever have been shown to exhibit increased total immunoglobulin E (IgE) levels in blood. IgE is also known as the reagenic antibody. In general, elevated levels of IgE indicate an increased probability of an IgE-mediated hypersensitivity, responsible for allergic reactions. Parasitic infestations such as hookworm, and certain clinical disorders including aspergillosis, have also been demonstrated to cause high levels of IgE. Decreased levels of IgE are found in cases of hypogammaglobulinemia, autoimmune diseases, ulcerative colitis, hepatitis, cancer, and malaria. Cord blood or serum IgE levels may have prognostic value in assessing the risk of future allergic conditions in children.

Certain groups of white blood cells, including basophils and tissue mast cells, have membrane receptors for the IgE molecule. These target cells, through a series of complex reactions, form a combination of a specific

allergen with antibody-sensitized basophils such as histamine, into the blood stream. As a result of these biochemical mediators, there is a constriction of smooth muscles, dilation of small blood vessels, activation of blood platelets, and irritation of skin nerve endings characteristic of allergic reactions. Typical clinical symptoms of immediate hypersensitivity are inflammation and itching in a skin reaction, or congestion in a bronchial reaction.

The IgE serum concentration in a patient is dependent on both the extent of the allergic reaction and the number of different allergens to which he is sensitized. Nonallergic normal individuals have IgE concentrations that vary widely and increase steadily during childhood, reaching their highest levels at age 15 to 20, and thereafter remaining constant until about age 60, when they slowly decline.

The IgE Quantitative Enzyme Immunoassay provides a rapid, sensitive, and reliable assay for total serum IgE. The minimal sensitivity of this assay is about 5.0 IU/ml.

#### Principle of the test

The IgE Quantitative Test Kit is based on a solid phase enzyme-linked immunosorbent assay. The assay system utilizes one anti-IgE antibody for solid phase (microtiter wells) immobilization and another anti-IgE antibody in the antibody-enzyme (horseradish peroxidase) conjugate solution. The test specimen (serum) is added to the IgE antibody coated microtiterwells and incubated with the Zero Buffer. If human IgE is present in the specimen, it will combine with the antibody on the well. The well is then washed to remove any residual test specimen, and IgE antibody labeled with horseradish peroxidase (conjugate) are added. The conjugate will bind immunologically to the IgE on the well, resulting in the IgE molecules being sandwiched between the solid phase and enzyme-linked antibodies. After a incubation at room temperature, the wells are washed with water to remove unbound labeled antibodies. A solution of TMB is added and incubated for 20 minutes, resulting in the development of a blue color. The color development is stopped with the addition of 2N HCl, and the color is changed to yellow and measured spectrophotometrically at 450 nm. The concentration of IgE is directly proportional to the color intensity of the test sample.

#### Materials and components

##### **Materials provided with the test kits:**

- Antibody-coated microtiter wells, 96 wells per plate.
- Reference standards, 0, 10, 50, 100, 400, and 800 IU/ml. Liquid, ready for use.
- Zero Buffer, 12 ml.
- Enzyme Conjugate Reagent, 18 ml.
- TMB Substrate, 12 ml.

- Stop Solution, 12 ml.
- Wash Buffer Concentrate(50X),15ml
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**Materials required but not provided:**

- Precision pipettes: 10µl~40µl,40µl~200µl and 1.0ml.
- Disposable pipette tips.
- Distilled water.
- Vortex mixer or equivalent.
- Absorbent paper or paper towel.
- Graph paper.
- Microtiter well reader.

**Specimen collection and preparation**

Serum should be prepared from a whole blood specimen obtained by acceptable medical techniques. This kit is for use with serum samples without additives only.

**Storage of test kits and instrumentation**

Unopened test kits should be stored at 2-8°C upon receipt and the microtiter plate should be kept in a sealed bag with desiccants to minimize exposure to damp air. Opened test kits will remain stable until the expiring date shown, provided it is stored as prescribed above. A microtiter plate reader with a bandwidth of 10nm or less and an optical density range of 0-2 OD or greater at 450nm wavelength is acceptable for use in absorbance measurement.

**Reagent preparation**

- All reagent should be brought to room temperature (18-22°C ) before use.
- Dilute 1 volume of Wash Buffer (50x) with 49 volumes of distilled water. For example, Dilute 15 ml of Wash Buffer (50x) into distilled water to prepare 750 ml of washing buffer (1x). Mix well before use.

**Assay procedures**

1. Secure the desired number of coated wells in the holder.
2. Dispense 20µl of standard, specimens, and controls into appropriate wells.
3. Dispense 100µl of Zero Buffer into each well.
4. Thoroughly mix for 10 seconds. It is very important to have complete mixing in this setup.
5. Incubate at room temperature (18-22°C) for 30 minutes.
6. Remove the incubation mixture by flicking plate content into a waste container.
7. Rinse and flick the microtiter wells 5 times with washing buffer (1X).

8. Strike the wells sharply onto absorbent paper or paper towels to remove all residual water droplets.
9. Dispense 150µl of Enzyme Conjugate Reagent into each well. Gently mix for 5 seconds.
10. Incubate at room temperature for 30 minutes.
11. Remove the incubation mixture by flicking plate contents into sink.
12. Rinse and flick the microtiter wells 5 times with washing buffer (1X).
13. Strike the wells sharply onto absorbent paper or paper towels to remove all residual water droplets.
14. Dispense 100µl TMB solution into each well. Gently mix for 5 seconds.
15. Incubate at room temperature in the dark for 20 minutes.
16. Stop the reaction by adding 100µl of Stop Solution to each well.
17. Gently mix for 30 seconds. It is important to make sure that all the blue color changes to yellow color completely.
18. Read optical density at 450nm with a microtiter reader.

**Important Note:**

The wash procedure is critical. Insufficient washing will result in poor precision and falsely elevated absorbance readings.

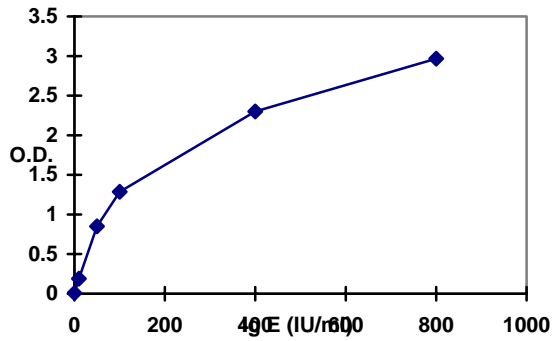
**Calculation of results**

Calculate the mean absorbance value (A<sub>450</sub>) for each set of reference standards, specimens, controls and patient samples. Constructed a standard curve by plotting the mean absorbance obtained from each reference standard against its concentration in IU/ml on graph paper, with absorbance values on the vertical or Y axis and concentrations on the horizontal or X axis. Use the mean absorbance values for each specimen to determine the corresponding concentration of IgE in IU/ml from the standard curve.

**Example of standard curve**

Results of typical standard run with optical density reading at 450nm shown in the Y axis against IgE concentrations shown in the X axis. This standard curve is for the purpose of illustration only, and should not be used to calculate unknowns. Each user should obtain his or her own data and standard curve.

IgE (IU/ml)	Absorbance (450nm)
0	0.008
10	0.189
50	0.851
100	1.287
400	2.300
800	2.966



### Expected values and sensitivity

The total IgE level in a normal, allergy-free adult is less than 150 IU/ml of serum. The minimum detectable concentration of IgE by this assay is estimated to be 5.0 IU/ml.

### References

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2. Buckley R.H. **Immunopharmacology of Allergic Disease** 1979; 117
3. Michel F. B., Bousquet J. and Greilier P. **J. Allergy Clin. Immunol.** 1980; 64: 422
4. Ishizaka T. **Ann Allergy** 1982; 48: 313
5. Kulczynski A. Jr. **J. Allergy Clin. Immunol.** 1981; 68: 5

Date Adopted	Reference No.
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