

AccuDiag™ Cashew ELISA Kit

REF 5158-8



Test	Cashew ELISA
Recovery	93 - 109%
Incubation Time	60 min
Sensitivity (Cashew)	0.2 ppm

INTENDED USE

The Cashew tree (*Anacardium occidentale*) belongs to the family of Anacardiaceae. With about 18% the fraction of proteins in cashew seed is very high. Some of these proteins, like the vicilin Ana o 1, the legumin Ana o 2 or the albumin Ana o 3 are known for being allergenic. Many of them are heat resistant making them stable to different production processes. For this reason cashew represents an important food allergen.

For cashew-allergic persons hidden cashew allergens in food are a critical problem. Already very low amounts of cashew can cause allergic reactions, which may lead to anaphylactic shock in severe cases. Because of this, cashew-allergic persons must strictly avoid the consumption of cashew containing food. Cross-contamination, mostly in consequence of the production process, is often noticed. This explains why in many cases the existence of cashew residues in food cannot be excluded. For this reason sensitive detection systems for cashew residues in foodstuffs are required.

The DAI Cashew ELISA represents a highly sensitive detection system for cashew and is particularly capable of the quantification of residues in cookies, cereals, ice-cream and chocolate.

TEST PRINCIPLE

The DAI Cashew quantitative ELISA test is based on the principle of the enzyme linked immunosorbent assay. An antibody directed against cashew proteins is bound on the surface of a microtiter plate. Cashew containing samples or standards are given into the wells of the microtiter plate. After 20 minutes incubation at room temperature, the wells are washed with diluted washing solution to remove unbound material. A peroxidase conjugated second antibody directed against cashew proteins is given into the wells and after 20 minutes of incubation the plate is washed again. A substrate solution is added and incubated for 20 minutes, resulting in the development of a blue color. The color development is terminated by the addition of a stop solution, and the color turns yellow. The yellow color is measured photometrically at 450 nm. The concentration of cashew is directly proportional to the color intensity of the test sample.

MATERIALS AND COMPONENTS

Materials provided with the test kits

The kit contains reagents for 96 determinations. They have to be stored at 2-8°C. Expiry data are found on the labels of the bottles and the outer package.

1. Microtiter plate consisting of 12 strips with 8 breakable wells each, coated with anti-cashew antibodies.
2. Cashew Standards (0, 2, 6, 20, 60 ppm of cashew): 5 vials with 4.0 mL each, dyed red, ready-to-use.
3. Conjugate (anti-cashew-peroxidase): 15 mL, dyed red, ready-to-use.
4. Substrate Solution (TMB): 15 mL, ready-to-use.
5. Stop Solution (0.5 M H₂SO₄): 15 mL, ready-to-use.
6. Extraction and sample dilution buffer (Tris): 2 x 120 mL as 10x concentrate, dyed red. Dilute 1+9 with distilled water. Stored at 4°C the diluted buffer is stable for at least one week. If during the cold storage crystals precipitate, the concentrate should be warmed up to 37°C for 15 minutes.
7. Washing Solution (PBS + Tween 20): 60 mL as 10x concentrate. Dilute 1+9 with distilled water. Stored at 4°C the diluted buffer is stable for at least 4 weeks. If during the cold storage crystals precipitate, the concentrate should be warmed up to 37°C for 15 minutes.
8. Plastic bag to store unused microtiter strips.
9. Instruction Manual.

Materials required but not provided

Instrumentation

1. 100 - 1000 µL micropipettes
2. Volumetric flask
3. Analytical balance
4. Mortar, mixer
5. Water bath
6. Centrifuge
7. ELISA reader (450 nm)

Reagents

Double-distilled water

SAMPLE PREPARATION

Due to the high risk of cross-contamination all applied instruments like applicator, mortar, glass vials etc. have to be **cleaned thoroughly** before and after each sample. To identify possible cross-contamination caused by previous extractions it is strongly recommended to note the sequence of the extractions.

The following sample preparation should be applied for **solid** samples:

1. To maximize homogeneity and representative-ness of the sample drawing, a minimum of 5 g sample should be pulverized finely in a mortar, impact mill, etc.
2. 1 g of the homogenized mixture is suspended in 20 mL of pre-diluted extraction and sample dilution buffer. Afterwards the suspension is incubated for 15 min in a preheated water bath at 60°C. To ensure good homogeneity, the samples should be shaken every two minutes.
3. The samples are centrifuged for 10 minutes at 2000 g. If it is not possible to separate the supernatant from the precipitate completely, the suspension should be filtrated if necessary.
4. 100 µL of particle-free solution are applied per well. If the results of a sample are out of the measuring range, further dilution with the pre-diluted extraction and sample dilution buffer is necessary. The additional dilution has to be considered when calculating the concentration.

The following sample preparation should be applied for **liquid** samples:

1 mL of liquid sample is diluted in 19 mL of pre-diluted extraction and sample dilution buffer. Afterwards the suspension is incubated for 15 min in a preheated water bath at 60°C. To ensure good homogeneity, the samples should be shaken every two minutes. The process is continued at point 3 of solid sample extraction process.

PRECAUTION

Full compliance of the following good laboratory practices (GLP) will determine the reliability of the results:

1. Prior to beginning the assay procedure, bring all reagents to room temperature (20-25°C).
2. All reagents should be mixed by gentle inversion or swirling prior to use. Do not induce foaming.
3. Once the assay has been started, all subsequent steps should be completed without interruption and within the recommended time limits.
4. Replace caps in all the reagents immediately after use. Do not interchange vial stoppers.
5. Use a separate disposable tip for each specimen to prevent cross-contamination.
6. All specimens and standards should be run at the same time, so that all conditions of testing are the same.
7. Do not mix components from different batches.
8. Do not use reagents after expiration date.
9. Check both precision and accuracy of the laboratory equipment used during the procedure (micropipettes, ELISA reader etc.).

HEALTH & SAFETY INSTRUCTIONS

1. Do not smoke or eat or drink or pipet by mouth in the laboratory.
2. Wear disposable gloves whenever handling patient specimens.
3. Avoid contact of substrate and stop solution with skin and mucosa (possible irritation, burn or toxicity hazard). In case of contact, rinse the affected zone with plenty of water.
4. Handling and disposal of chemical products must be done according to good laboratory practices (GLP).

ASSAY PROCEDURE

The washing solution is supplied as 10x concentrate and has to be **diluted** 1+9 with double distilled water before use.

In any case the **ready-to-use** standards should be determined twofold. When samples in great quantities are determined, the standards should be pipetted once before the samples and once after the samples. For final interpretation the arithmetic mean is used for calculation.

In consideration of GLP and quality control requirements a duplicate measurement of samples is recommended.

The procedure is according to the following scheme:

1. Prepare samples as described above.
2. Pipet 100 µL ready-to-use standards or prepared samples in duplicate into the appropriate wells of the microtiter plate.
3. Incubate for 20 minutes at room temperature.
4. Wash the plate three times as follows: Discard the contents of the wells (dump or aspirate). Pipet 300 µL of diluted washing solution into each well. After the

third repetition empty the wells again and remove residual liquid by striking the plate against a paper towel. The wash procedure is critical. Insufficient washing will result in poor precision and falsely elevated absorbencies.

5. Pipet 100 µL of conjugate (anti-cashew-peroxidase) into each well.
6. Incubate for 20 minutes at room temperature.
7. Wash the plate as outlined in 4.
8. Pipet 100 µL of substrate solution into each well.
9. Allow the reaction to develop in the dark (e.g. cupboard or drawer; the chromogen is light-sensitive) for 20 minutes at room temperature.
10. Stop enzyme reaction by adding 100 µL of stop solution (0.5 M H₂SO₄) into each well. The blue color will turn yellow upon addition.
11. After thorough mixing, measure absorbance at 450 nm (reference wavelength 620 nm), using an ELISA reader. The color is stable for 30 minutes.

RESULTS

The ready-to-use standards are prepared for a direct determination of sample concentrations. The dilution of samples in the extraction process as described in the above stated sample preparation procedure is already considered. Additional dilution due to high sample concentration has to be accounted for.

1. Calculate the average optical density (OD 450 nm) for each set of reference standards or samples.
2. Construct a standard curve by plotting the mean optical density obtained for each reference standard against its concentration in ppm on semi-log graph paper with the optical density on the vertical (y) axis and the concentration on the horizontal (x) axis. Alternatively the evaluation can be carried out by software. In this case the 4-parameter method should be preferred.
3. Using the mean optical density value for each sample, determine the corresponding concentration of cashew in ppm from the standard curve. Depending on experience and/or the availability of computer capability, other methods of data reduction may be employed.

Typical Standard Values

The following table contains an example for a typical standard curve. The OD% is calculated as percent of the absorption of the 60 ppm standard. These values are only an example and should not be used instead of the standard curve which has to be measured in each new test.

Cashew (ppm)	OD% of 60 ppm
60	100
20	50
6	18
2	7
0	1

PERFORMANCE CHARACTERISTICS

Sensitivity

The limit of detection (LOD) of the **DAI Cashew ELISA** test is 0.2 ppm for the standard curve.

Validation experiments with common matrices resulted in the following LODs (ppm).

Cookies	0.0
Cereals	0.3
Ice cream	0.3
Dark chocolate	0.5

The limit of quantification (LOQ) of the **DAI Cashew ELISA** test is 2 ppm.

Due to the variety of sample matrices and their influence on the blank, results less than the LOQ should be treated as negative.

Cross-reactivity

For the following foods no cross-reactivity could be detected:

Almond	Cow's milk	Pork
Apricot	Egg	Potato
Barley	Gliadin	Pumpkin seed
Bean	Goat's milk	Rice
Beef	Guar gum	Rye
Bovine gelatin	Hazelnut	Sucrose
Brazil nut	Kiwi	Sesame
Buckwheat	Lentil	Sheep's milk
Carob gum	Lupine	Shrimps, cooked
Carrot	Macadamia	Shrimps, raw
Celery	Mustard	Soy
Chestnut	Oats	Soy lecithin
Chicken	Pea	Sunflower seed
Chickpea	Peanut	Tomato
Cocoa	Pecan	Walnut
Coconut	Pine seed	Wheat
Cod	Plum	
Corn	Poppy seed	

The following cross-reactions were determined:

Pistachio	4%
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Precision

Intra-assay Precision	4-5%
Inter-assay Precision	3-6%

Linearity

The serial dilution of spiked samples (cookies, cereals, ice cream, and dark chocolate) resulted in a dilution linearity of 107%-131%.

Recovery

Mean recovery was determined by spiking samples with different amounts of cashew:

Cookies	109%
Cereals	98%
Ice cream	93%
Dark chocolate	102%

REFERENCES

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