OneStep Leptospira IgG/IgM RapiCard™ InstaTest (Serum/Plasma/Whole Blood)

TEST PRINCIPLE

The Leptospira IgG/IgM RapiCard™ (Whole Blood/Serum/Plasma) is a qualitative, membrane based immunoassay for the detection of IgG and IgM antibodies to Leptospira in whole blood, serum or plasma. The membrane is coated with recombinant Leptospira antibodies on the test line region of the cassette. During testing, the whole blood, serum or plasma specimen reacts with recombinant Leptospira antigens conjugated colloidal gold. The mixture migrates upward on the membrane chromatographically by capillary action to react with mouse anti-human IgG or mouse anti-human IgM on the membrane and generate a colored line. Presence of this colored line indicates a positive result, whereas its absence indicates a negative result. To serve as a procedural control, a colored line will always appear at the control line region indicating that proper volume of specimen has been added and membrane wicking has occurred.

SPECIMEN COLLECTION AND PREPARATION

- The Leptospira IgG/IgM RapiCard™ (Whole Blood/Serum/Plasma) can be performed using whole blood (from venipuncture or fingerstick), serum or plasma.
- To collect Fingerstick Whole Blood specimens:
  - Wash the patient’s hand with soap and warm water or clean with an alcohol swab. Allow to dry.
  - Massage the hand without touching the puncture site by rubbing down the hand towards the fingertip of the middle or ring finger.
  - Puncture the skin with a sterile lancet. Wipe away the first sign of blood.
  - Gently rub the hand from wrist to palm to form a rounded drop of blood over the puncture site.
  - Add the Fingerstick Whole Blood specimen to the test by using a capillary tube:
    - Touch the end of the capillary tube to the blood until filled to approximately 40 μL. Avoid air bubbles.
    - Place the bulb onto the top end of the capillary tube, then squeeze the bulb to dispense the whole blood to the specimen area of the test cassette.
  - Add the Fingerstick Whole Blood specimen to the test by using hanging drops:
    - Position the patient’s finger so that the drop of blood is just above the specimen area of the test cassette.
    - Allow 1 hanging drop of fingerstick whole blood to fall into the center of the specimen area on the test cassette, or move the patient’s finger so that the hanging drop touches the center of the specimen area. Avoid touching the finger directly to the specimen area.
    - Separate the serum or plasma from blood as soon as possible to avoid hemolysis. Only clear, non-hemolyzed specimens can be used.
    - Testing should be performed immediately after the specimens have been collected. Do not leave the specimens at room temperature for prolonged periods. Serum and plasma specimens may be stored at 2-8°C for up to 3 days. For long term storage, specimens should be kept below 20°C. Whole blood collected by venipuncture should be stored at 2-8°C if the test is to be run within 2 days of collection. Do not freeze whole blood specimens. Whole blood collected by fingerstick should be tested immediately.
- Bring specimens to room temperature prior to testing. Frozen specimens must be completely thawed and mixed well prior to testing. Specimens should not be frozen and thawed repeatedly.
- If specimens are to be shipped, they should be packed in compliance with federal regulations for transportation of etiologic agents.

REAGENTS

The test cassette contains recombinant Leptospira antigens conjugated colloidal gold, mouse anti-human IgG and mouse anti-human IgM coated on the membrane.

Materials provided
- Test cassettes
- Droppers
- Buffer

Materials required but not provided
- Specimen collection containers
- Centrifuge (for plasma only)
- Timer
- Lancets (for fingerstick whole blood only)
- Heparinized capillary tubes and dispensing bulb (for fingerstick whole blood only)
ASSAY PROCEDURE

Allow test cassette, specimen, and/or controls to equilibrate to room temperature (15-30°C) prior to testing.
1. Bring the pouch to room temperature before opening it. Remove the test cassette from the sealed pouch and use it as soon as possible. Best results will be obtained if the assay is performed within one hour.
2. Place the cassette on a clean and level surface. For Serum or Plasma specimen: Hold the dropper vertically and transfer 1 drop of serum or plasma (approximately 40 μL) to the specimen area, then add 2 drops of buffer (approximately 80 μL) and start the timer, see illustration below. For Venipuncture Whole Blood specimen: Hold the dropper vertically and transfer 1 drop of whole blood (approximately 40 μL) to the specimen area, then add 2 drops of buffer (approximately 80 μL) and start the timer. See illustration below. For Fingerstick Whole Blood specimen:
   • To use a capillary tube: Fill the capillary tube and transfer approximately 40 μL of fingerstick whole blood specimen to the specimen area of test cassette, then add 2 drops of buffer (approximately 80 μL) and start the timer. See illustration below.
   • To use hanging drops: Allow 1 hanging drop of fingerstick whole blood specimen (approximately 40 μL) to fall into the specimen area of test cassette, then add 2 drops of buffer (approximately 80 μL) and start the timer. See illustration below.
3. Wait for the colored line(s) to appear. Read the result at 15 minutes, do not interpret the result after 20 minutes.

RESULTS

(Please refer to the illustration above)

IgG POSITIVE: Two distinct colored lines appear. One color line should be in the control region (C) and another color line should be in the IgG region.
IgM POSITIVE: Two distinct colored lines appear. One color line should be in the control region (C) and another color line should be in the IgM region.
IgG and IgM POSITIVE: Three distinct colored lines appear. One color line should be in the control region (C) and another two color lines should be in the IgG and IgM region.
NOTE: The intensity of the color in the test line region (T) will vary depending on the concentration of Leptospira antibodies present in the specimen. Therefore, any shade of red in the test region should be considered positive.
NEGATIVE: One color line appears in the control region (C). No apparent red or pink line appears in the IgG and IgM region.
INVALID: Control line fails to appear. Insufficient specimen volume or incorrect procedural techniques are the most likely reasons for control line failure. Review the procedure and repeat the test with a new test cassette. If the problem persists, discontinue using the test kit immediately and contact your local distributor.

QUALITY CONTROL

Internal procedural controls are included in the test. A color line appearing in the control region (C) is an internal positive procedural control. It confirms sufficient specimen volume and correct procedural technique. Control standards are not supplied with this kit; however, it is recommended that positive and negative controls be tested as a good laboratory practice to confirm the test procedure and to verify proper test performance.

PRECAUTION

• For professional in vitro diagnostic use only. Do not use after expiration date.
• Do not eat, drink or smoke in the area where the specimens or kits are handled.

• Handle all specimens as if they contain infectious agents. Observe established precautions against microbiological hazards throughout the procedure and follow the standard procedures for proper disposal of specimens.
• Wear protective clothing such as laboratory coats, disposable gloves and eye protection when specimens are assayed.
• Humidity and temperature can adversely affect results.

STORAGE

The kit can be stored at room temperature or refrigerated (2-30°C). The test cassette is stable through the expiration date printed on the sealed pouch. The test cassette must remain in the sealed pouch until use. DO NOT FREEZE. Do not use beyond the expiration date.

LIMITATIONS

1. The Direction for Use and the Interpretation of Result must be closely when testing the presence of antibodies to pathogenic L. interrogans in serum, plasma or whole blood from individual subjects. Failure to follow the procedure may give inaccurate results.
2. The Leptospira IgG/IgM RapiCard™ (Whole Blood/Serum/Plasma) is limited to the qualitative detection of antibodies to Leptospira interrogans in human serum, plasma or whole blood. The intensity of the test band does not have a linear correlation with antibody titer in the specimen.
3. A negative result for an individual subject indicates absence of detectable Leptospira interrogans antibodies. However, a negative test result does not preclude the possibility of exposure to Leptospira interrogans.
4. A negative result can occur if the quantity of Leptospira interrogans antibodies present in the specimen is below the detection limits of the assay, or the antibodies that are detected are not present during the stage of disease in which a sample is collected.
5. Some specimens containing unusually high titer of heterophile antibodies or rheumatoid factor may affect expected results.
6. The results obtained with this test should only be interpreted in conjunction with other diagnostic procedures and clinical findings.
EXPECTED VALUES

The Leptospira IgG/IgM RapiCard™ (Whole Blood/Serum/Plasma) has been compared with a leading commercial Leptospira IgG/IgM EIA test. The correlation between these two systems is 98%.

PERFORMANCE CHARACTERISTICS

Sensitivity and Specificity

A total of 210 samples from susceptible subjects were tested by the Leptospira IgG/IgM RapiCard™ and by a commercial Leptospira IgM EIA kit. Comparison for all subjects is shown in the following table.

<table>
<thead>
<tr>
<th>Method</th>
<th>IgM Results</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Results</td>
</tr>
<tr>
<td>Leptospira IgG/IgM</td>
<td></td>
</tr>
<tr>
<td>RapiCard™ (Whole Blood/Serum/Plasma)</td>
<td>Positive</td>
</tr>
<tr>
<td></td>
<td>Negative</td>
</tr>
<tr>
<td>Total Result</td>
<td></td>
</tr>
</tbody>
</table>

Relative sensitivity: 90% (95% CI: 55.5–99.7%)
Relative specificity: 99.0% (95% CI: 96.4–99.9%)
Accuracy: 98.6% (95% CI: 95.9–99.7%)

*Confidence Intervals

A total of 206 samples from susceptible subjects were tested by the Leptospira IgG/IgM RapiCard™ and by a commercial Leptospira IgM EIA kit. Comparison for all subjects is shown in the following table.

<table>
<thead>
<tr>
<th>Method</th>
<th>IgG Results</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Results</td>
</tr>
<tr>
<td>Leptospira IgG/IgM</td>
<td></td>
</tr>
<tr>
<td>RapiCard™ (Whole Blood/Serum/Plasma)</td>
<td>Positive</td>
</tr>
<tr>
<td></td>
<td>Negative</td>
</tr>
<tr>
<td>Total Result</td>
<td></td>
</tr>
</tbody>
</table>

Relative sensitivity: > 99.9% (95% CI: 60.7%–100%)
Relative specificity: 99.0% (95% CI: 96.4–99.9%)
Accuracy: 99.0% (95% CI: 96.5%–99.9%)

*Confidence Intervals

Within-run precision has been determined by using 20 replicates of three specimens: a negative, a Leptospira IgM low titer positive, a Leptospira IgM high titer positive, a Leptospira IgG low titer positive and a Leptospira IgG high titer positive. The negative, a Leptospira IgM low titer positive, a Leptospira IgM high titer positive, a Leptospira IgG low titer positive and a Leptospira IgG high titer positive values were correctly identified 100% of the time.

Inter-Assay

Between-run precision has been determined by 20 independent assays on the same three specimens: a negative, a Leptospira IgM low titer positive, a Leptospira IgM high titer positive, a Leptospira IgG low titer positive and a Leptospira IgG high titer positive. Three different lots of the Leptospira IgG/IgM RapiCard™ (Whole Blood/Serum/Plasma) have been tested over a 3-month period using negative, a Leptospira IgM low titer positive, a Leptospira IgG high titer positive and a Leptospira IgG high titer positive specimens. The specimens were correctly identified 100% of the time.

Cross-reactivity

The Leptospira IgG/IgM RapiCard™ (Whole Blood/Serum/Plasma) has been tested by HAMA, RF, HBsAg, HBsAb, HBeAg, HBCab, Syphilis, HIV, H. Pylori, MONO, CMV, Rubella and TOXO positive specimens. The results showed no cross-reactivity.

Interfering Substances

The following potentially interfering substances were added to HCV negative and positive specimens.

Acetaminophen: 20 mg/dL
Caffeine: 20 mg/dL
Acetylsalicylic Acid: 20 mg/dL
Genistic Acid: 20 mg/dL
Ascorbic Acid: 2g/dL
Albumin: 2 g/dL
Creatin: 200 mg/dL
Hemoglobin 1000mg/dL
Bilirubin: 1g/dL
Oxalic Acid: 60mg/dL

None of the substances at the concentration tested interfered in the assay.

REFERENCE


Diagnostic Automation/ Cortez Diagnostics, Inc.
21250 Califa St, Suite 102 and 116, Woodland Hills, California 91367 USA

Date Adopted: 2016-06-21

CEpartner4U, Esdoornlaan 13, 3951DB Maarn. The Netherlands. www.cepartner4u.eu

Revision Date: 2015-09-08